

Analysis of Mixtrue Experiment data set

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Introduction

This data set was generated by Aliaksei Holik from Asselin-Labat Laboratory at the WEHI. FASTQ files from this experiment are available from GEO under accession number GSE64098. The matrix of counts analysed below was obtained by running Subread and featureCounts from the Rsubread package. The data set is from mixture experiment with total RNA libraries mapped to the reverse strand infomation. The R code below generated the results presented in Liu et al. (2014).

Importing the data

```
library(edgeR)
library(limma)
load("mixtureExperiment.rda")
counts= tstrand$counts
Repl=as.factor(tstrand$samples[, "Replicate"])
group=as.factor(tstrand$samples[, "group"])

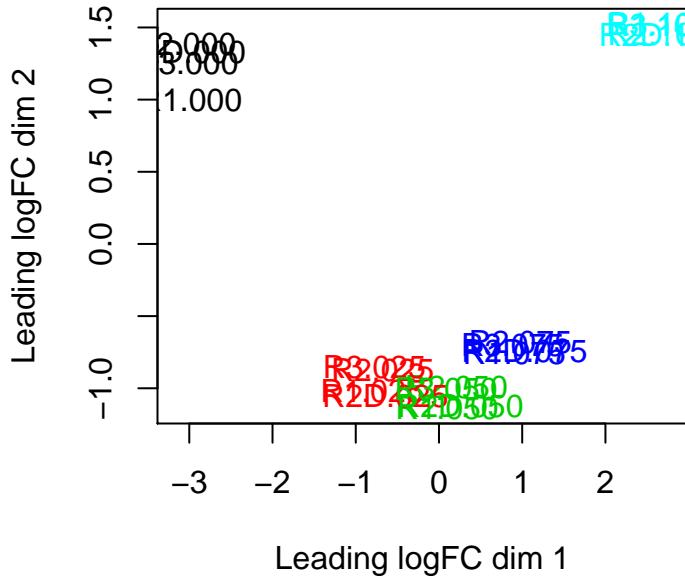
des=model.matrix(~0+group)
colnames(des) = c("mix0", "mix25", "mix50", "mix75", "mix100")

x=calcNormFactors(tstrand, method="TMM")
goodrep2 = 6:10
dodgyrep2 = 11:15

des2 = model.matrix(~tstrand$samples$group[-dodgyrep2]-1)
colnames(des2) = c("mix0", "mix25", "mix50", "mix75", "mix100")

#seperate data into different groups #
RSCE.good = calcNormFactors(tstrand[,-dodgyrep2])
RSCE.dodgy = calcNormFactors(tstrand[,-goodrep2])
RSCE.remove=calcNormFactors(tstrand[,-c(6:15)])

# Make an MDS plot: include all dodgy and good samples
plotMDS(x, labels=tstrand$sample$Sample, col=as.numeric(tstrand$sample$group))
```



Voom analysis

```
# Analysis of good and dodgy samples using voom only on full data set
vgoodonly=voom(RSCE.good,des2,plot=F)
vdodgyonly=voom(RSCE.dodgy,des2,plot=F)
contr = makeContrasts("100vs0"=mix100-mix0,"75vs0"=mix75-mix0,
                      "50vs0"=mix50-mix0,"25vs0"=mix25-mix0,levels=des2)
fitgoodonly = lmFit(vgoodonly)
cfitgoodonly = contrasts.fit(fitgoodonly, contr)
cfitgoodonly = eBayes(cfitgoodonly)

fitdodgyonly = lmFit(vdodgyonly)
cfitdodgyonly = contrasts.fit(fitdodgyonly, contr)
cfitdodgyonly = eBayes(cfitdodgyonly)

dtgoodonly = decideTests(cfitgoodonly)
dtdodgyonly = decideTests(cfitdodgyonly)

summary(dtdodgyonly)

##      100vs0 75vs0 50vs0 25vs0
## -1     5631   4522   2965    558
## 0     11741  13452  16220  20665
## 1      5772   5170   3959   1921
```

```

summary(dtgoodonly)

##      100vs0 75vs0 50vs0 25vs0
## -1    6037  5112  3720  1053
## 0    10882 12371 14932 19714
## 1     6225  5661  4492  2377

```

Voom with sample weights

```

# Analysis of Degraded samples using combined voom and sample quality weights
#figure 8A
vdodgya = voomWithQualityWeights(RSCE.dodgy, des2, plot=F)

contr = makeContrasts("100vs0"=mix100-mix0,"75vs0"=mix75-mix0,
                      "50vs0"=mix50-mix0,"25vs0"=mix25-mix0,levels=des2)

fitdodgya = lmFit(vdodgya)
cfitdodgya = contrasts.fit(fitdodgya, contr)
cfitdodgya = eBayes(cfitdodgya)

dtdodgya = decideTests(cfitdodgya)

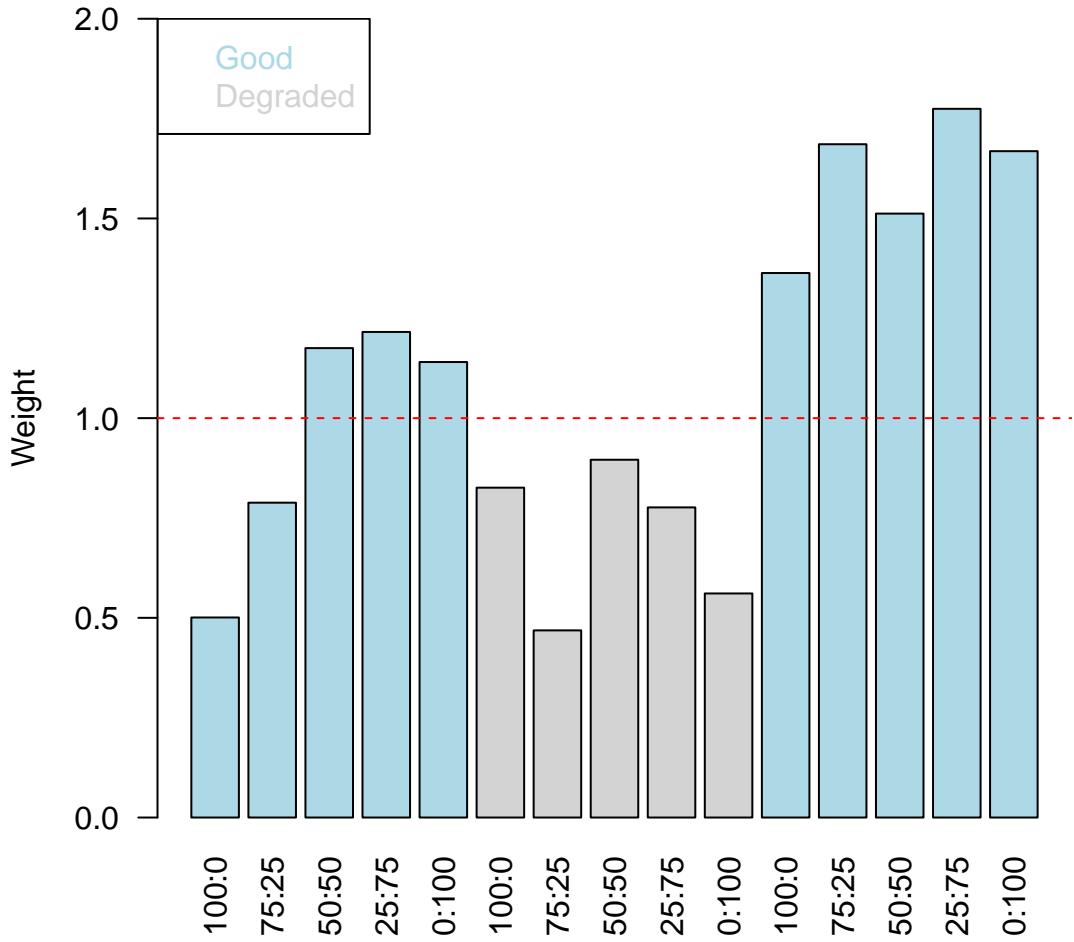
summary(dtdodgya)

##      100vs0 75vs0 50vs0 25vs0
## -1    5895  4849  3250   589
## 0    11272 12894 15703 20620
## 1     5977  5401  4191  1935

barplot(vdodgya$sample, main="Figure 8(A)", col=rep(c("lightblue", "lightgrey", "lightblue"),
                                                 times=c(5,5,5)), ylab="Weight",
         names=rep(c("100:0", "75:25", "50:50", "25:75", "0:100"), times=3), las=2, ylim=c(0,2))
legend("topleft", legend=c("Good", "Degraded"), text.col=c("lightblue", "lightgrey"))
abline(h=1, col=2, lty=2)

```

Figure 8(A)



Voom after removing outlier samples

```
# Analysis where outlier samples have been removed
des3=model.matrix(~tstrand$samples$group[-c(6:15)]-1)
colnames(des3) = c("mix0", "mix25", "mix50", "mix75", "mix100")

vremove = voom(RSCE.remove, des3, plot=F)

contrRemove = makeContrasts("100vs0"=mix100-mix0,"75vs0"=mix75-mix0,
                            "50vs0"=mix50-mix0,"25vs0"=mix25-mix0,levels=des3)
fitremove = lmFit(vremove)
cfitremove = contrasts.fit(fitremove, contrRemove)
```

```
cfitremove = eBayes(cfitremove)

dtremove = decideTests(cfitremove)

summary(dtremove)

##    100vs0 75vs0 50vs0 25vs0
## -1    5231  4042  2413   393
## 0    12610 14552 17494 21480
## 1     5303  4550  3237  1271
```

Voom with block weights

```
# Analysis of Degraded samples using combined voom and blocked sample quality weights
#figure 8B
Z <- cbind(c(1,1,1,1,1,0,0,0,0,1,1,1,1,1),c(0,0,0,0,0,1,1,1,1,1,0,0,0,0))

vdodgyb = voomWithQualityWeights(RSCE.dodgy, design=des2,
  normalization="none", var.design=Z, plot=F)
contr = makeContrasts("100vs0"=mix100-mix0,"75vs0"=mix75-mix0,
  "50vs0"=mix50-mix0,"25vs0"=mix25-mix0,levels=des2)

fitdodgyb = lmFit(vdodgyb)
cfitdodgyb = contrasts.fit(fitdodgyb, contr)
cfitdodgyb = eBayes(cfitdodgyb)

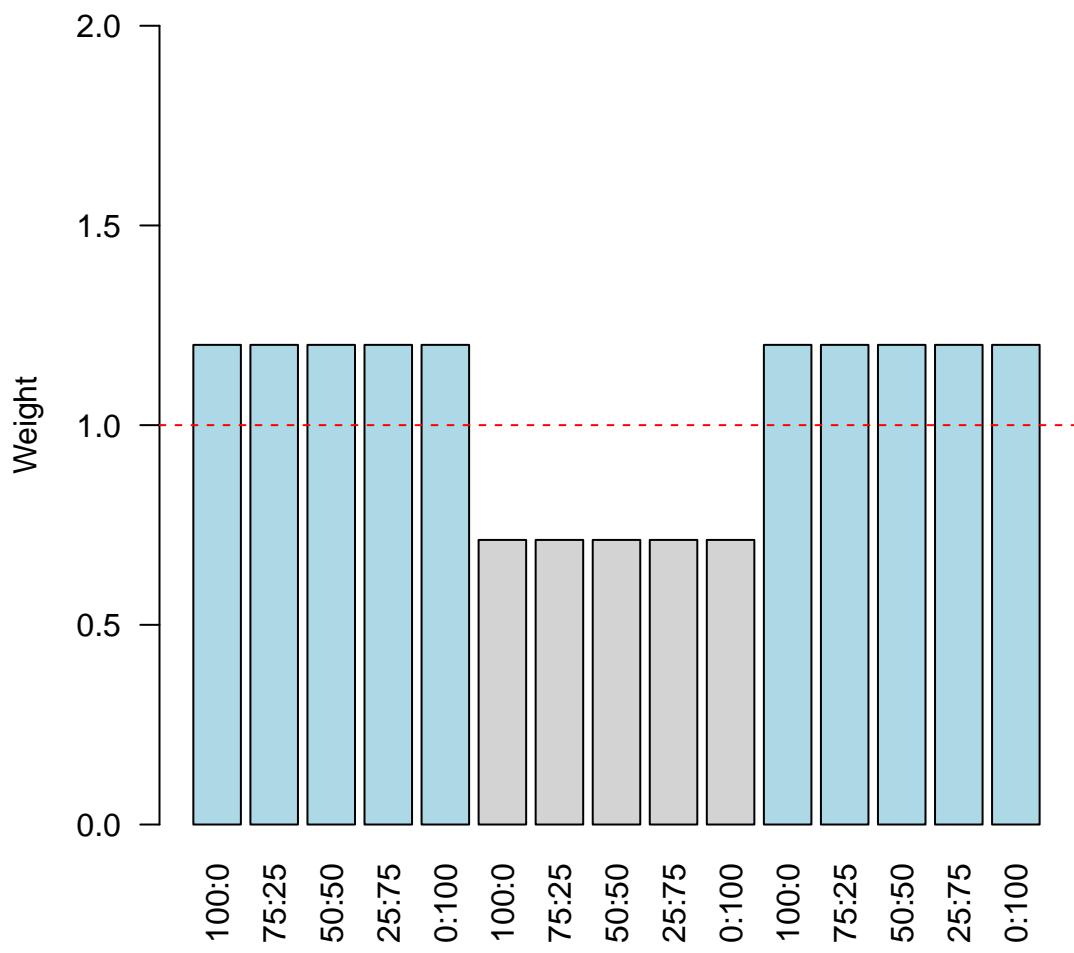
dtdodgyb = decideTests(cfitdodgyb)

summary(dtdodgyb)

##      100vs0 75vs0 50vs0 25vs0
## -1    5760   4673   3090   622
## 0    11482  13208  15992  20540
## 1     5902   5263   4062   1982

barplot(vdodgyb$sample, main="Figure 8(B)", col=rep(c("lightblue", "lightgrey", "lightblue"),
  times=c(5,5,5)), ylab="Weight",
  names=rep(c("100:0", "75:25", "50:50", "25:75", "0:100"), times=3), las=2, ylim=c(0,2))
abline(h=1, col=2, lty=2)
```

Figure 8(B)



Overlap DE genes between good samples and four methods

```
#Checking the agreement between DE genes from analysis of good sample and DE genes from analysis with dodgy samples#
n=n1=matrix(NA,4,5)
for(i in 1:4){
coegoodonly=cfitgoodonly$genes$GeneID[dtgoodonly[,i] !=0]
coedodgonly=cfitdodgonly$genes$GeneID[dtdodgonly[,i] !=0]
coedodgya=cfitdodgya$genes$GeneID[dtdodgya[,i] !=0]
coedodgby=cfitdodgby$genes$GeneID[dtdodgby[,i] !=0]
coerremove=cfitremove$genes$GeneID[dtremove[,i] !=0]

n[i,]=c(length(intersect(coegoodonly,coedodgonly)),
length(intersect(coegoodonly,coedodgya)),length(intersect(coegoodonly,coedodgby)),
length(intersect(coegoodonly,coerremove)),length(coegoodonly))
n[i,]=n[i,]/length(coegoodonly)*100

n1[i,]=c(length(coedodgonly),length(coedodgya),length(coedodgby),length(coerremove),length(coegoodonly))

}
n=apply(n,2,function(x){paste(round(x,1), "%", sep="")})

colnames(n)=c("Degraded-Voom", "Dodgy-Voom+Sample",
"Degraded-voom+block", "Remove Outliers", "Good analysis")
colnames(n1)=c("Degraded-Voom", "Dodgy-Voom+Sample",
"Degraded-voom+block", "Remove Outliers", "Good analysis")
rownames(n)=rownames(n1)=c("100vs0", "75vs0", "50vs0", "25vs0")
n

##          Degraded-Voom Dodgy-Voom+Sample Degraded-voom+block
## 100vs0 "88.7%"      "90.4%"      "90.4%"
## 75vs0  "84.8%"      "87.2%"      "86.8%"
## 50vs0  "78.1%"      "81.5%"      "80.8%"
## 25vs0  "64%"        "64.6%"      "67.3%"
##          Remove Outliers Good analysis
## 100vs0 "82.7%"      "100%"
## 75vs0  "76.6%"      "100%"
## 50vs0  "65.6%"      "100%"
## 25vs0  "44.8%"      "100%"

n1

##          Degraded-Voom Dodgy-Voom+Sample Degraded-voom+block
## 100vs0      11403         11872         11662
## 75vs0       9692          10250         9936
## 50vs0       6924          7441          7152
## 25vs0       2479          2524          2604
##          Remove Outliers Good analysis
```

```
## 100vs0      10534      12262
## 75vs0       8592       10773
## 50vs0       5650       8212
## 25vs0       1664       3430
```

```
sessionInfo()

## R version 3.1.1 (2014-07-10)
## Platform: x86_64-unknown-linux-gnu (64-bit)
##
## locale:
## [1] LC_CTYPE=en_AU.UTF-8      LC_NUMERIC=C
## [3] LC_TIME=en_AU.UTF-8       LC_COLLATE=en_AU.UTF-8
## [5] LC_MONETARY=en_AU.UTF-8   LC_MESSAGES=en_AU.UTF-8
## [7] LC_PAPER=en_AU.UTF-8     LC_NAME=C
## [9] LC_ADDRESS=C              LC_TELEPHONE=C
## [11] LC_MEASUREMENT=en_AU.UTF-8 LC_IDENTIFICATION=C
##
## attached base packages:
## [1] stats      graphics   grDevices utils      datasets  methods
## [7] base
##
## other attached packages:
## [1] edgeR_3.8.2  knitr_1.8    limma_3.23.1
## 
## loaded via a namespace (and not attached):
## [1] evaluate_0.5.5 formatR_1.0   highr_0.4    stringr_0.6.2
## [5] tools_3.1.1
```